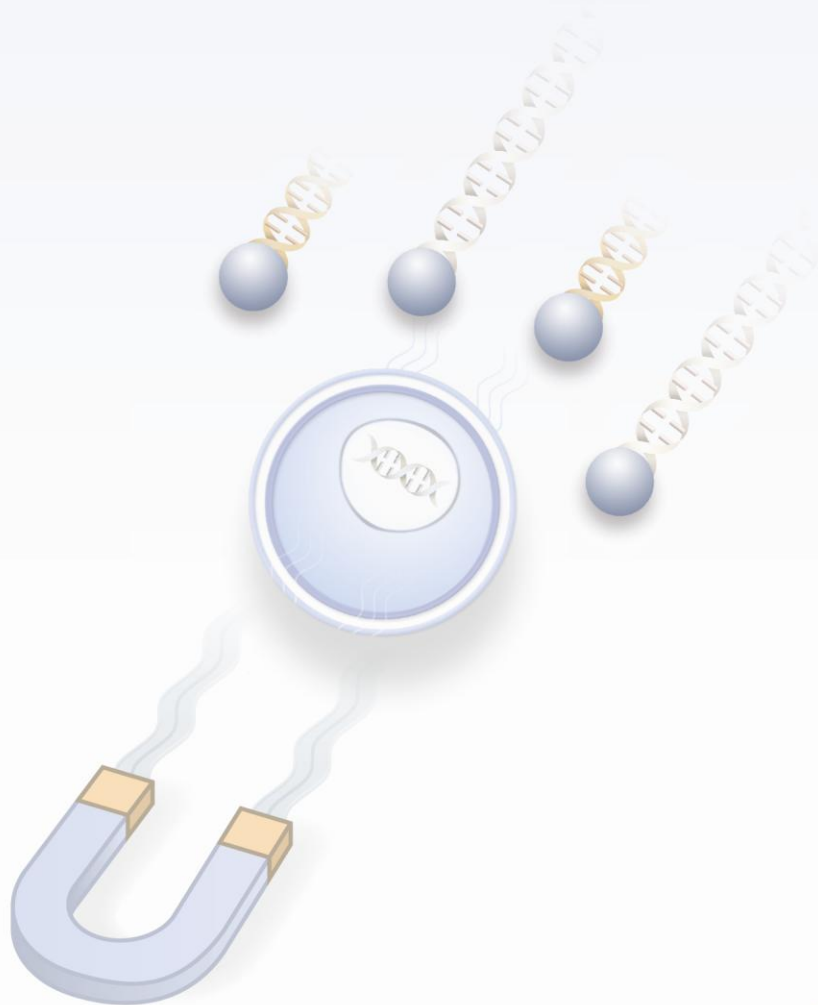


Magnetofectamine™

Transfection Kit:

Lipofectamine™ 2000 + CombiMag



Magnetofectamine™

Instruction Manual

The two most powerful transfection technologies in one kit!

Magnetofectamine™ is the association of the world-known **Lipofectamine™ 2000** transfection reagent from Invitrogen™, with the unique Magnetofection™ based reagent **CombiMag** from OZ Biosciences™

List of Magnetofectamine™ kits

Catalog Number	Description	Volume (µL)	Number of transfection per µg of DNA
MTX0750	Magnetofectamine™ Kit ¹	250µL + 750µL	250-375
MTX1000	Magnetofectamine™ Starting Kit ²	250µL + 750µL	250-375

¹ Contains 1 vial of each reagent (250µL of CombiMag + 750µL of Lipofectamine™ 2000)

² Contains 1 vial of each reagent and a Magnetic plate (250µL of CombiMag + 750µL of Lipofectamine™ 2000 + Super Magnetic plate MF10000)

Use the content of the table above to determine the appropriate catalog number for your needs. You can order these products by contacting us (order@ozbiosciences.com). For all other supplementary information, do not hesitate to contact our dedicated technical support (tech@ozbiosciences.com).

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1. Technology

1.1. Description

OZ Biosciences is thrilled to announce the association of its universal Magnetofection-based reagent, **CombiMag**, with **Lipofectamine™ 2000** the most renowned transfection reagent from Invitrogen™ in one convenient kit: **Magnetofectamine™**.

The alliance of **Lipofectamine™ 2000** and **CombiMag** reagent leads to increased transfection efficiency, minimized toxicity and enhanced gene expression. It is specially suited for hard-to-transfect and primary cells.

Magnetofection™ is a creative, simple and highly efficient method to transfect cells *in vitro* and *in vivo* *. This magnetic assisted transfection technique is based on the association of magnetic nanoparticles with nucleic acids (alone or pre-complexed with a transfection reagent) or viruses to which a magnetic field is exerted. The magnetic force drives the gene vector towards the target cells, allows the vector dose to get concentrated onto the cell very rapidly and triggers delivery via endocytosis. Consequently, high transfection efficiencies can be achieved with less nucleic acid amount.

Lipofectamine™ 2000 transfection reagent is a proprietary formulation of Invitrogen™ for transfecting nucleic acids (DNA and siRNA) into eukaryotic cells.

Magnetofectamine™ main advantages:

1. Improved transfection efficiency
2. Less nucleic acids used - minimized toxicity
3. Ideal for hard-to-transfect and primary cells
4. No need to change your standard Lipofectamine™ 2000 protocol
5. Compatible with any types of nucleic acid and any culture medium
6. Simple, ready-to-use & rapid

* Plank C, Zelphati O, and Mykhaylyk O. Magnetically enhanced nucleic acid delivery. Ten years of magnetofection-Progress and prospects. *Adv Drug Deliv Rev.* 2011; 63 (14-15):1300-31.

Contents

- The kit: CombiMag (250µL) and Lipofectamine™ 2000 (750µL) suitable for 250-375 transfections/µg of DNA.
- The Starting kit: the Magnetofectamine™ kit (see above) and a Super Magnetic Plate.

Stability, Storage and Shipping

Storage: Upon reception, store the CombiMag reagent and the Lipofectamine™ 2000 reagent at +4°C.

- **Do not freeze the magnetic nanoparticles!**
- **Do not add anything to the stock solution of magnetic nanoparticles!**
- **Do not freeze Lipofectamine™ 2000 transfection reagent!**

Stability: Magnetofectamine™ kits are stable for at least 18 months at the recommended storage temperature.

Shipping condition: Magnetofectamine™ kit is shipped on wet ice.

2. Applications

Magnetofectamine™ Kit associates Lipofectamine™ 2000 transfection reagent and CombiMag, the magnetic nanoparticles designed to be combined with any transfection reagents and to boost their efficiencies. The combination of the two technologies enables using smaller amounts of nucleic acid and increasing the overall efficiency of your transfection. For further information concerning the Magnetofection™ technology, see our website: www.ozbiosciences.com.

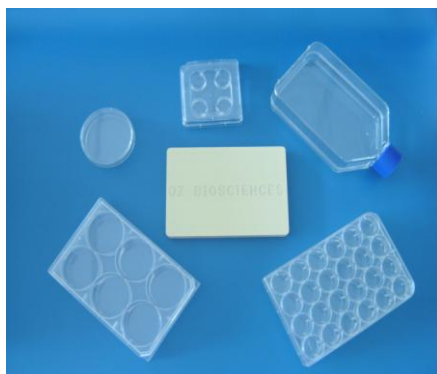
Magnetofectamine™ Kit can be used with various types of nucleic acids (DNA, mRNA, siRNA...). It is suitable for numerous cell types and is particularly useful for very difficult-to-transfect cells & primary cells. An updated list of successfully transfected cells is available on OZ Biosciences website: www.ozbiosciences.com. You can also submit your data to tech@ozbiosciences.com so we can update this list and give you all the support you need. Magnetofectamine™ Kit is serum compatible and can be used for transient and stable transfection. These products are very stable, ready-to-use and intended for research purpose only (not intended for diagnostic use or any animal and human therapeutic).

3. Magnetofection™ Apparatus

As for all Magnetofection™ reagents, Magnetofectamine™ Kit requires appropriate magnetic fields. Magnetic plates especially designed for Magnetofection are provided to exert specific magnetic fields. Their special geometry produces strong magnetic fields that are suitable for all cell culture dishes (T-75 flasks, 60 & 100 mm dishes, 6-, 12-, 24-, 48- and 96-well plates). Two standard magnetic plates: 96-magnet plate and Super Magnetic plate, and one Mega Magnetic plate are available. Super Magnetic plate suits for all cell culture supports and Mega Magnetic plate is designed to hold up to 4 culture dishes once. The magnetic plate design allows producing a heterogeneous magnetic field that magnetizes the nanoparticles in solution, forms a very strong gradient and covers all the surface of the plate.



Magnetic plate 96 magnets



Super Magnetic Plate



Mega Magnetic Plate

4. Protocol

4.1. General Considerations / Important Guidelines

The instructions given below represent sample protocols that were successfully applied with a variety of cells. Optimal conditions may vary depending on the nucleic acid, cell types, size of cell culture dishes and presence or absence of serum. Therefore, the amounts and ratio of the individual components may have to be adjusted to achieve best results. As a starting point, use **2 or 3 μL of Lipofectamine™ 2000 and 1 μL of CombiMag per μg of DNA.** It is recommended to transfect cells at high cell density for high efficiency, high expression levels, and to minimize cytotoxicity.

- **Cells** should be healthy and assayed during their exponential growing phase. The presence of contaminants (mycoplasma, fungi) will considerably affect the transfection efficiency. The optimal confluency has to be adjusted according to the cells used. We recommend using regularly passaged cells for transfection. Do not use cells that have been cultured for too long (> 2 months).
- **Nucleic acid** should be as pure as possible. Endotoxins levels must be very low since they interfere with transfection efficiencies. We suggest avoiding long incubation time of the diluted DNA/RNA solution before the reagents addition to circumvent any degradation or surface adsorption. For plasmid DNA use the protocol in section 4.3 and for siRNA transfection use the protocol in section 4.4.
- **Culture Medium.** Presence of antibiotics in the culture medium during the transfection is not recommended. Opti-MEM is suggested to dilute the Lipofectamine™ 2000 and nucleic acids.

4.2. Cells Preparation

Adherent cells. It is recommended to plate the cells the day prior transfection in growth medium without antibiotics. Cells should be **90-95 % confluent** at the time of transfection (see the suggested cell number in the Table 1). The correct choice of optimal plating density also depends on the planned time between transfection and transgene analysis: for a large interval, we recommend a lower density and for a short interval a higher density may be advantageous.

Suspension cells, co-transfection (DNA/DNA or DNA/siRNA) and reverse transfection protocols are also available. Please contact our technical support department (tech@ozbiosciences.com) for those specific procedures.

Table 1: Cell number suggested.

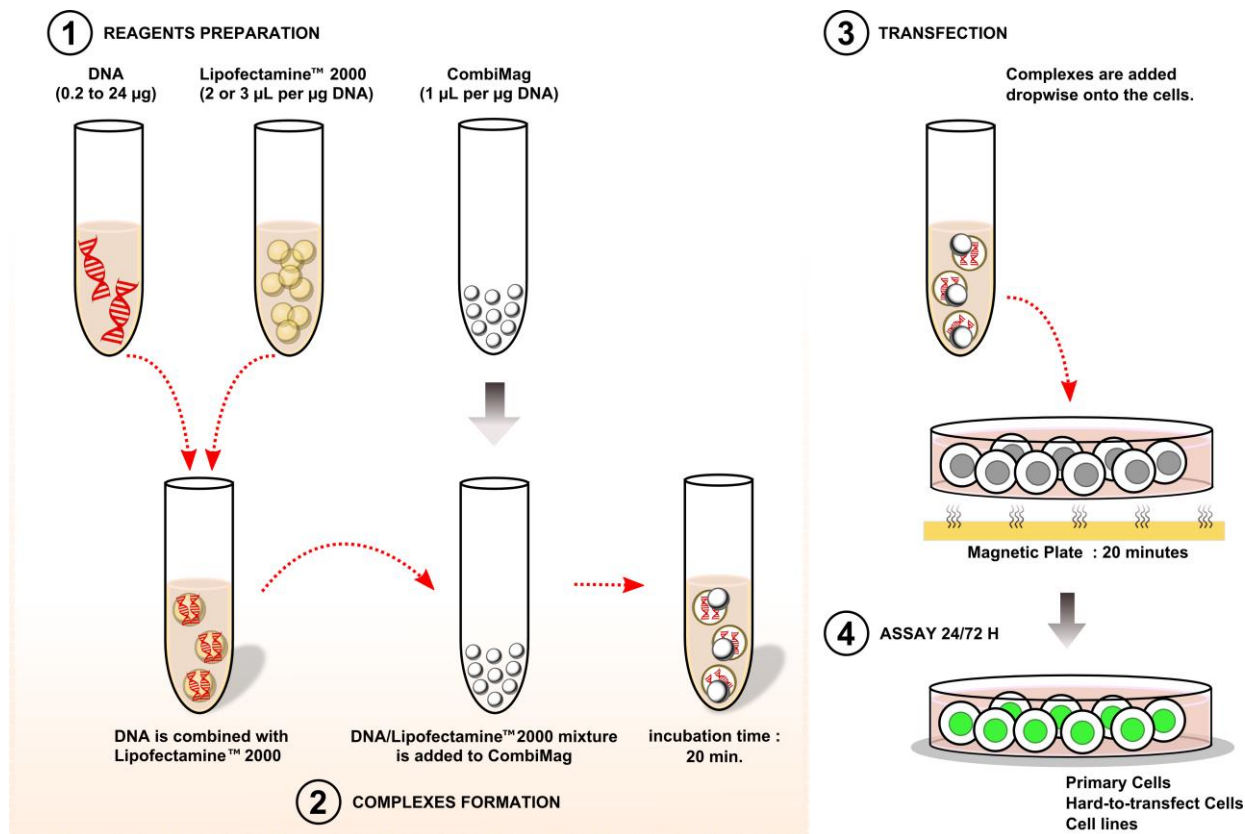
Tissue Culture Dish	Surface area per well ¹	Cell Number
96 well	0.3 cm ²	0.05 – 0.2 x 10 ⁵
24 well	2 cm ²	0.5 – 2 x 10 ⁵
12 well	4 cm ²	1 – 4 x 10 ⁵
6 well	10 cm ²	2 – 10 x 10 ⁵
60 mm	20 cm ²	5 – 20 x 10 ⁵
100 mm	60 cm ²	10 – 60 x 10 ⁵

¹ Surfaces area may vary depending on the manufacturer.

4.3. DNA Transfection Protocol

Table 2 shows transfection conditions according to different cell culture formats (all amounts are given on per-well basis). The DNA, Lipofectamine™ 2000 and CombiMag solutions should be at room temperature and be gently vortexed prior to use.

The rapid protocol, suitable for most cell lines, is as simple as follows: use **2 or 3 μL of Lipofectamine™ 2000 and 1 μL of CombiMag per μg of DNA.** We suggest beginning with this ratio and optimize it, if required.



1) **Cells.** One day before transfection prepare the cells as indicated in 4.2

2) **Reagents preparation**

- DNA solution.** Dilute 0.2 to 24 µg of DNA in 25 to 1500 µL of Opti-MEM® I Reduced Serum Medium or culture medium without serum (see Table 2).
- Lipofectamine™ 2000 solution.** Gently mix the reagent before use. Dilute 0.5 to 60 µL of Lipofectamine™ 2000 in 25 to 1500 µL of Opti-MEM® I Reduced Serum Medium or culture medium without serum (see Table 2). Incubate for 5 minutes at room temperature.
- CombiMag reagent.** Vortex the reagent before each use. Use 1 µL CombiMag / µg DNA (see Table 2). Add the CombiMag reagent directly into a tube (do not dilute with any culture medium).

Note: For DNA doses of less than 1 µg pre-dilute an aliquot of CombiMag reagent with deionized water only and use the volume required for your DNA dose. Discard the diluted CombiMag after use.

Note: Proceed to step 3 within 10 minutes.

Table 2: DNA amount, Lipofectamine™ 2000 volume and transfection conditions suggested.

Tissue Culture Dish	DNA Quantity (µg)	Lipofectamine™ 2000 Volume (µL)	Dilution Volume (µL) ¹	CombiMag Volume (µL)	Total culture medium Volume
96 well	0.2	0.5	2 x 25	0.2	100 µL
24 well	0.8	2	2 x 50	0.8	500 µL
12 well	1.6	4	2 x 100	1.6	1 mL
6 well	4.0	10	2 x 250	4.0	2 mL
60 mm	8.0	20	2 x 500	8.0	5 mL
100 mm	24.0	60	2 x 1500	24.0	15 mL

¹ Volumes of dilution medium in step 2a and 2b.

3) **Complexes formation.** Combine the DNA solution with the Lipofectamine™ 2000 solution. Mix gently and add the DNA/Lipofectamine™ 2000 mixture immediately to the CombiMag Reagent. Mix gently and incubate for 20 minutes at room temperature.

Note: Proceed to step 4 within 30 minutes

4) **Transfection.**

- a. Add the complexes prepared previously dropwise onto the cells growing in serum-containing culture medium and homogenize by rocking the plate back and forth.
- b. Incubate the cells 20 minutes on the magnetic plate at room temperature or at 37°C in a CO₂ incubator. After incubation remove the magnetic plate.

Optional: Right after this step, you may perform a change of medium. Keep the magnetic plate beneath the cell culture dish, withdraw the old medium and add fresh growth medium (optionally without antibiotics). Then, remove the magnetic plate.

- 5) **Assay.** Incubate the cells at 37°C in a CO₂ incubator under standard conditions until evaluation of the transgene expression. Depending on the cell type and promoter activity, the assay for the reporter gene can be performed 24 to 72 hours following transfection.

Note: For some cells, 24 hours post-transfection replace the old media with fresh media or just add fresh growth culture medium to the cells.

Note: In the case of cells very sensitive to transfection, the medium can be changed after 4-6 hours of incubation with fresh medium.

Stable transfection. The same protocol can be used to produce stably transduced cells except that 48 or 72 hours post-transfection, passage the cells at a 1:10 (or higher dilution) into fresh medium containing the appropriate antibiotics for selection.

Other protocols for suspension cells, reverse transfection or co-transfection (DNA/DNA or DNA/siRNA) are available on our website at www.ozbiosciences.com or by contacting our technical support department (tech@ozbiosciences.com).

Optimization protocol for DNA transfection.

To achieve the highest transfection efficiency, optimize the transfection conditions as follows:

- Vary the DNA (µg) /Lipofectamine™ 2000 (µL) ratio from 1/0.5 to 1/5.
- Once the optimal DNA/Lipofectamine™ 2000 ratio is found, adjust the CombiMag quantity with 0.5, 1 and 2 µL of CombiMag per µg of DNA.
- Thereafter, with the optimal ratios previously found, adjust the DNA quantity according to Table 3.
- Finally, culture medium composition (for preparing the complexes), cell density, total culture medium volume and incubation times can also be optimized.

Table 3: Suggested range of DNA amounts for optimization.

Tissue Culture Dish	DNA Quantity (µg)
96 well	0.1 to 0.5
24 well	0.5 to 2
12 well	1 to 4
6 well	2 to 10
60 mm dish	5 to 30
100 mm dish	15 to 60

4.4. siRNA Transfection Protocol

Table 4 shows transfection conditions according to different cell culture formats (all amount are given on per-well basis). The siRNA, Lipofectamine™ 2000 and CombiMag solutions should be at room temperature and be gently vortexed prior to use.

The rapid protocol, suitable for most cell lines, is as simple as follows: **use 2 µL of Lipofectamine™ 2000 and 1 µL of CombiMag with 40 pmol of siRNA.** We suggest beginning with this ratio and optimize it, if required.

- 1) **Cells.** The day prior transfection, prepare the cells in growth medium without antibiotics such as they will be 30-50% confluent at the time of transfection.

Note: Transfecting cells at a lower density allows a longer interval between transfection and assay time, and minimizes the loss of cell viability due to cell overgrowth.

2) Reagents preparation

- siRNA solution.* Dilute 5 to 600 pmol of siRNA in 25 to 1500 μL of Opti-MEM[®] I Reduced Serum Medium or culture medium without serum (see Table 4). Mix gently. The final concentration of siRNA when added to the cells is 33nM.
- Lipofectamine™ 2000 solution.* Gently mix the reagent before use. Dilute 0.25 to 30 μL of Lipofectamine™ 2000 in 25 to 1500 μL of Opti-MEM[®] I Reduced Serum Medium or culture medium without serum (see Table 4). Incubate for 5 minutes at room temperature.
- CombiMag reagent.* Vortex the reagent before each use. Use 1 μL of CombiMag for 40 pmol of siRNA (or for 2 μL of Lipofectamine™ 2000; see Table 4). Add the CombiMag reagent directly into an empty tube (do not dilute with any culture medium).

Note: For siRNA doses of less than 40 pmol pre-dilute an aliquot of CombiMag reagent with deionized water only and use the volume required for your siRNA dose. Discard the diluted CombiMag after use.

Note: Proceed to step 3 within 10 minutes.

Table 4: siRNA amount, Lipofectamine™ 2000 volume and transfection conditions suggested.

Tissue Culture Dish	siRNA Quantity (pmol)	Lipofectamine™ 2000 Volume (μL)	Dilution Volume (μL) ¹	CombiMag Volume (μL)	Total culture medium Volume
96 well	5	0.25	2 x 25	0.125	100 μL
24 well	20	1	2 x 50	0.5	500 μL
12 well	40	2	2 x 100	1	1 mL
6 well	100	5	2 x 250	2.5	2.5 mL
60 mm	200	10	2 x 500	5	5 mL
100 mm	600	30	2 x 1500	15	15 mL

¹ Volumes of dilution medium in step 2a and 2b.

- Complexes formation.** Combine the siRNA solution with the Lipofectamine™ 2000 solution. Mix gently and add the siRNA/Lipofectamine™ 2000 mixture immediately to the CombiMag Reagent. Mix gently and incubate for 20 minutes at room temperature.

Note: Proceed to step 4 within 30 minutes

4) Transfection.

- Add the complexes prepared previously dropwise onto the cells growing in serum-containing culture medium and homogenize by rocking the plate to ensure a uniform distribution of the mixture.
- Incubate the cells 20 minutes on the magnetic plate at room temperature or at 37°C in a CO₂ incubator. After incubation remove the magnetic plate.

Optional: Right after this step, you may perform a change of medium. Keep the magnetic plate beneath the cell culture dish, withdraw the old medium and add fresh growth medium (optionally without antibiotics). Then, remove the magnetic plate.

- Assay.** Incubate the cells at 37°C in a CO₂ incubator for 24 to 96 hours until evaluation of the gene silencing.

Note: For some cells, the medium can be changed after 4-6 hours of incubation with fresh medium.

Note: For the most powerful gene knockdown a second sequential transfection (24 hours later) can be realized under the same conditions.

Other protocols for suspension cells, reverse transfection or co-transfection (DNA/siRNA) are also available on our website at www.ozbiosciences.com or by contacting our technical support department (tech@ozbiosciences.com).

Optimization protocol for siRNA transfection.

To achieve the highest transfection efficiency and minimize off target effects, optimize the transfection conditions as follows:

- Vary the siRNA (pmol) /Lipofectamine™ 2000 (µL) ratio from 40/1 to 40/5.
- Once the optimal siRNA/Lipofectamine™ 2000 ratio is found, adjust the CombiMag quantity with 0.5, 1 and 2 µL of CombiMag per 40 pmol of siRNA (or 2 µL of Lipofectamine™ 2000).
- Thereafter, with the optimal ratios previously found, adjust the siRNA quantity according to Table 5.
- Finally, culture medium composition (for preparing the complexes), cell density, total culture medium volume and incubation times can also be optimized.

Table 5: Suggested range of siRNA amounts for optimization.

Tissue Culture Dish	siRNA Quantity (pmol)
96 well	1.25 to 10
24 well	5 to 40
12 well	10 to 80
6 well	25 to 200
60 mm dish	50 to 400
100 mm dish	150 to 1200

OZ Biosciences team has developed a detail protocol for optimization and also cell specific optimal transfection procedure. Thus, do not hesitate to contact our technical service at tech@ozbiosciences.com to request these specific protocols.

5. Appendix

For any question related to Lipofectamine™ 2000 use, please contact directly the manufacturer. Our dedicated and specialized technical support team will be pleased to answer any of your requests and to help you with your experiments at tech@ozbiosciences.com. In addition, do not hesitate to visit our website www.ozbiosciences.com and the FAQ section.

5.1 Quality Controls

To assure the performance of each lot of CombiMag produced, we qualify each component using rigorous standards. The following *in vitro* assays are conducted to qualify the function, quality and activity of each kit component.

Specification	Standard Quality Controls
<i>Sterility</i>	Thioglycolate assay. Absence of fungal and bacterial contamination shall be obtained for 7 days.
<i>Biological Activity</i>	Transfection efficiency. Every lot shall have an acceptance specification of > 80% of the activity of the reference lot.
<i>Magnetic nanoparticles</i>	1. Quality and size homogeneity of the magnetic nanoparticles. 2. Stability of the magnetic nanoparticles formulations.

Lipofectamine™ 2000 is manufactured by Life Technologies Corporation for OZ Biosciences. Quality control of the Lipofectamine™ 2000 transfection reagent is also performed by Life Technologies Corporation according to their specifications, quality management system and certificate of analysis.

Problems	Comments and Suggestions
Low transfection efficiency	<p>1- Optimize Lipofectamine™ 2000/nucleic acid ratio. See section 4.3 and 4.4</p> <p>2- CombiMag / nucleic acid ratio. Optimize the lipoplexes / CombiMag ratio by varying the volume of CombiMag from 0.5 µL to 2 µL per µg of DNA or per 40 pmol of siRNA.</p> <p>3- DNA or siRNA amount. Use different quantity of DNA or siRNA with the recommended or optimized (above) transfection reagent / nucleic acid ratio.</p> <p>4- Cell density. A non-optimal cell density at the time of transfection can lead to insufficient uptake. The optimal confluency should be around 90% for DNA and 50% for siRNA but favorable cell density may vary according to the cell type. A mid-log growth phase is recommended.</p> <p>5- DNA or siRNA quality. Nucleic acids should be as pure as possible and free of contaminants (proteins, phenol, ethanol etc.). Endotoxins levels must be very low since they interfere with transfection efficiency. Use nuclease-free materials.</p> <p>6- Type of promoter. Ensure that DNA promoter can be recognized by the cells to be transfected. Another cell or viral-driven reporter gene expression can be used as a control.</p> <p>7- Cell condition. 1) Cells that have been in culture for a long time (> 8 weeks) may become resistant to transfection. Use freshly thawed cells that have been passaged at least once. 2) Cells should be healthy and assayed during their exponential growing phase. The presence of contaminants (mycoplasma, fungi) alters considerably the transfection efficiency.</p> <p>8- Medium used for preparing DNA or siRNA / transfection reagent complexes. It is critical that serum-free medium or buffer (HBS, PBS) are used during the preparation of the complexes.</p> <p>9- Cell culture medium composition. 1) For some cells, transfection efficiency can be increased using culture medium without serum or under reduced serum condition. In this case, we recommend transfecting cells in serum-free medium during the first 4h of incubation. 2) Presence of antibiotics might affect cell health and transfection efficiency.</p> <p>10- Incubation time and transfection volume. 1) Optimal time range between transfection and assay varies with cells, promoter, expression product, etc. Transfection efficiency can be monitored after 24 – 96h by analyzing the gene product. Several reporter genes can be used to quantitatively monitored gene expression kinetics. 2) To increase transfection efficiency, transfection volume suggested can be reduced for the first 24 hours.</p> <p>11- Old transfection reagent / DNA complexes. The transfection reagent / DNA complexes must be freshly prepared every time. Complexes prepared and stored for longer than 2 hours can be aggregated.</p> <p>12- Transgene detection assay. Ensure that your post-transfection assay is properly set up and includes a positive control.</p> <p>13- Transfection reagent temperature. Reagents have to be at room temperature and be vortexed prior to use.</p>
Cellular toxicity	<p>1- Unhealthy cells. 1) Check cells for contamination, 2) Use new batch of cells, 3) Ensure culture medium condition (pH, type of medium used, contamination etc), 4) Cells are too confluent or cell density is too low, 5) Verify equipments and materials</p> <p>2- Transgene product is toxic. Use suitable controls such as cells alone, transfection reagent alone or mock transfection with a DNA or siRNA control.</p> <p>3- siRNA/DNA quality - Presence of contaminants. Ensure that nucleic acid is pure, contaminant-free and endotoxin-free. Use high quality nucleic acids as impurities can lead to cell death.</p> <p>4- Concentration of transfection reagent / nucleic acid too high. Decrease the amount of nucleic acid / reagent complexes added to the cells by lowering the nucleic acid amount or the transfection reagent concentration. Complexes aggregation can cause some toxicity; prepare them freshly and adjust the ratio as outlined previously.</p>

	<p>5- Incubation time. Reduce the incubation time of complexes with the cells by replacing the transfection medium by fresh medium after 4h to 24h.</p> <p>6- Key gene silencing. If the targeted gene is essential for cell survival or if a key gene is non-specifically silenced by the siRNA, this can lead to cell death.</p>
No or weak gene silencing effect	<p>1- siRNA design. The design of an efficient siRNA is a crucial step. Ensure to use a validated siRNA sequence. If a validated siRNA cannot be used, assay your sequence in an easy to transfect cell line (if possible) in order to validate it (HeLa cells for example).</p> <p>2- siRNA concentration. Use higher amount of siRNA.</p> <p>3- Incubation time. Perform a time-course experiment to set up the optimal incubation time since gene silencing is dependent on the gene expression and the protein turnover rate.</p> <p>4- Medium used for preparing transfection reagent/siRNA complexes. It is critical that serum-free medium or buffer (HBS, PBS) are used during the complexes preparation.</p>

Our dedicated and specialized technical support group will be pleased to answer any of your requests and to help you with your transfection experiments: tech@ozbiosciences.com. In addition, do not hesitate to visit our website www.ozbiosciences.com and the FAQ section.

6. Related Products

Description
MAGNETOFECTION TECHNOLOGY
Super Magnetic Plate (<i>standard size for all cell culture support</i>) Mega Magnetic plate (<i>mega size to hold 4 culture dishes at one time</i>)
Transfection reagents:
PolyMag Neo (<i>for all nucleic acids</i>) / SilenceMag (<i>for siRNA application</i>) NeuroMag (<i>dedicated for neurons</i>)
Viral Transduction enhancers:
ViroMag (<i>to optimize viral transduction</i>) / AdenoMag (<i>for Adeno viruses</i>) ViroMag R/L (<i>specific for retrovirus and Lentivirus</i>)
LIPOFECTION TECHNOLOGY (LIPID-BASED)
Lullaby (<i>siRNA transfection reagent</i>) DreamFect Gold (<i>Transfection reagent for all types of nucleic acids</i>) VeroFect (<i>for Vero cells</i>) / FlyFectin (<i>for Insect cells</i>)
I-MICST TECHNOLOGY
Viro-MICST (<i>to transduce directly on magnetic cell purification columns</i>)
3D TRANSFECTION TECHNOLOGY
3Dfect (<i>for scaffolds culture</i>) / 3DfectIN (<i>for hydrogels culture</i>)
RECOMBINANT PROTEIN PRODUCTION
HYPE-5 Transfection Kit (<i>for High Yield Protein Expression</i>)
PROTEIN DELIVERY SYSTEMS
Ab-DeliverIN (<i>delivery reagent for antibodies</i>) Pro-DeliverIN (<i>delivery reagent for protein in vivo and in vitro</i>)
PLASMIDS PVECTOZ
pVectOZ-LacZ 25µg / pVectOZ-SEAP 25µg
ASSAY KITS
Bradford – Protein Assay Kit / MTT cell proliferation kit β-Galactosidase assay kits (CPRG/ONPG)
BIOCHEMICALS
D-Luciferin, K ⁺ and Na ⁺ 1g X-Gal powder 1g / G-418, Sulfate 1g

Do not hesitate to contact us for all complementary information and remember to visit our website in order to stay informed on our last breakthrough technologies and updated on our complete product list.

contact@ozbiosciences.com / www.ozbiosciences.com

Limited Use Label License for Lipofectamine™ 2000

This product contains Lipofectamine™ 2000 Transfection Reagent that is the subject of one or more issued U.S. patents or pending U.S. patent applications, and corresponding non-U.S. equivalents, owned by Life Technologies Corporation, and is sold under an agreement between Life Technologies Corporation and OZ Biosciences. The purchase price of this product includes limited, non-transferable rights under the foregoing patents to use this amount of the product only in research conducted by the buyer of the product. The buyer is not authorized to use this product or its components for any therapeutic, diagnostic or prophylactic purposes and is not authorized to use this product or its components for any commercial purpose. The buyer is not authorized to sell this product or its components, is not authorized to sell any products made using this product or its components, and is not authorized to perform a service with this product or its components. Further information about acquiring rights under patents owned by Life Technologies Corporation may be obtained by contacting the Licensing Department, Life Technologies Corporation, 5791 Van Allen Way, Carlsbad, CA 92008. (760) 603-7200. Email: outlicensing@lifetech.com

Limited License for Magnetofectamine™

The purchase of the Magnetofectamine™ Reagents grants the purchaser a non-transferable, non-exclusive license to use the kit and/or its separate and included components (as listed in section 1, Kit Contents). Reagents are intended **for in-house research only** by the buyer. Such use is limited to the transfection of nucleic acids as described in the product manual. In addition, research only use means that this kit and all of its contents are excluded, without limitation, from resale, repackaging, or use for the making or selling of any commercial product or service without the written approval of OZ Biosciences.

Separate licenses are available from OZ Biosciences for the express purpose of non-research use or applications of the Magnetofectamine™ Reagents. To inquire about such licenses, or to obtain authorization to transfer or use the enclosed material, contact the Director of Business Development at OZ Biosciences.

Buyers may end this License at any time by returning all Magnetofectamine reagent, material and documentation to OZ Biosciences, or by destroying all Magnetofectamine Kit components. Purchasers are advised to contact OZ Biosciences with the notification that a Magnetofectamine kit is being returned in order to be reimbursed and/or to definitely terminate a license for internal research use only granted through the purchase of the kit(s).

This document covers entirely the terms of Magnetofectamine reagent research only license, and does not grant any other express or implied license. The laws of the French Government shall govern the interpretation and enforcement of the terms of this License.

Product Use Limitations for Magnetofectamine™

The Magnetofectamine™ Reagents and all of its components are developed, designed, intended, and sold for research use only. They are not to be used for human diagnostic or included/used in any drug intended for human use or any therapeutic, diagnostic or prophylactic purposes and is not authorized to use this product or its components for any commercial purpose. . All care and attention should be exercised in the use of the kit components by following proper research laboratory practices. The buyer is not authorized to sell this product or its components, is not authorized to sell any products made using this product or its components, and is not authorized to perform a service with this product or its components.

For more information, or for any comments on the terms and conditions of this License, please contact:

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